

Native LESA mass spectrometry: Direct analysis of proteins and their complexes

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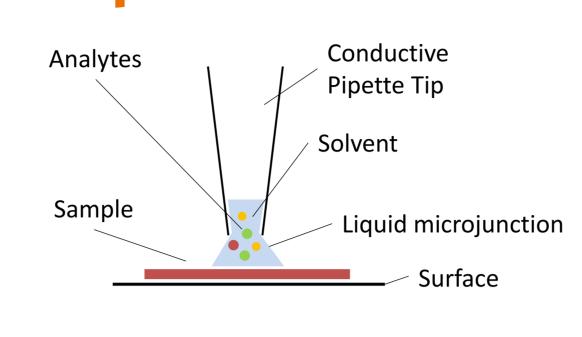
Overview

microjunction surface ammonium acetate based solvent systems is shown to be suitable for the analysis of a range of proteins in their native, folded, structure directly from complex sample substrates such as mouse liver and brain tissue.

Introduction

Liquid microjunction surface sampling via liquid extraction surface analysis (LESA) is an emerging tool for direct surface sampling of intact proteins and protein assemblies from biological substrates^{1,2,3}. Recently, ammonium acetate based buffers have been described as suitable sampling solvents^{4,5,6}. Here we show that native LESA MS can probe protein assemblies up to ~ 800 kDa and demonstrate improved methods for the analysis of folded intact protein species from thin tissue sections of bulk liver tissue and mouse brain for the first time. Furthermore, we show that chemical imaging of proteins and protein assemblies can be achived via native LESA MS. In addition, we explore native LESA MS for probing protein ligand-binding interactions from complex samples.

Experimental



Thin tissue sections (10 µm) thaw mounted onto glass slides were either analysed as is or after washing in 80 % ethanol. Human dried blood spots on filter paper were sampled a minimum of 24 hours after preparation. Protein standards were spotted onto

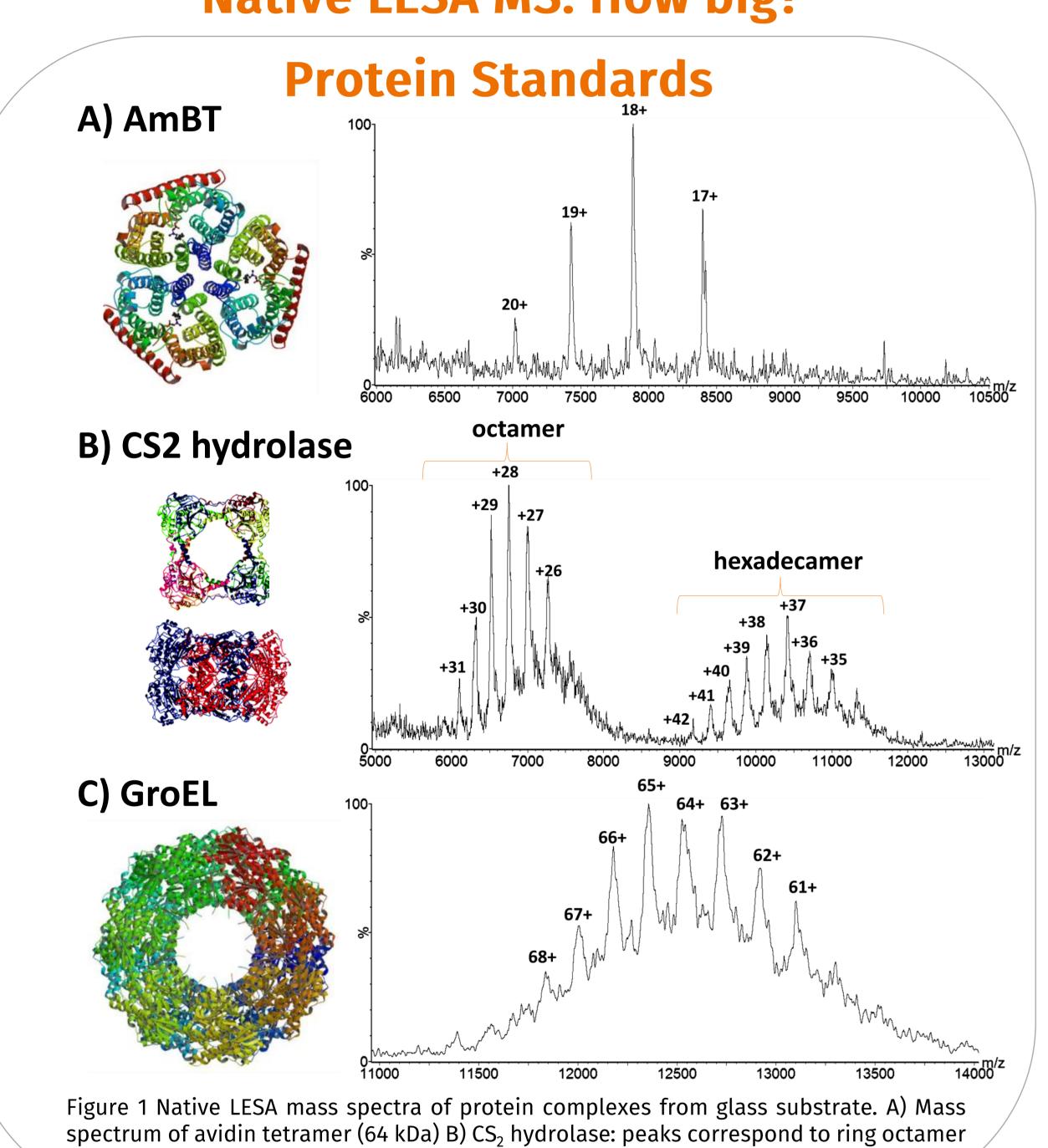
glass slides. A Triversa Nanomate (Advion™) was coupled to either a Synapt G2S mass spectrometer (Waters, UK) or an Orbitrap Elite mass spectrometer (Thermo Scientific, UK). LESA extraction solvent comprised 200 mM ammonium acetate + 5 % methanol, or the same solvent system containing 1 mM biotin or bezabibrate.

Native LESA MS: Imaging

Conclusions

- Native LESA MS is a suitable tool for direct analysis of intact proteins and protein assemblies from tissue substrates.
- Proteins can be spatially profiled using native solvent systems. Spatial distributions agree with previous reports and some proteins are described for the first time via LESA MS.
- Protein assemblies up to 800 kDa can be detected after extraction from a surface.
- High resolution native LESA data is shown on an Orbitrap Elite MS.

Native LESA MS: How big?



Native LESA MS: Thin Tissue Sections

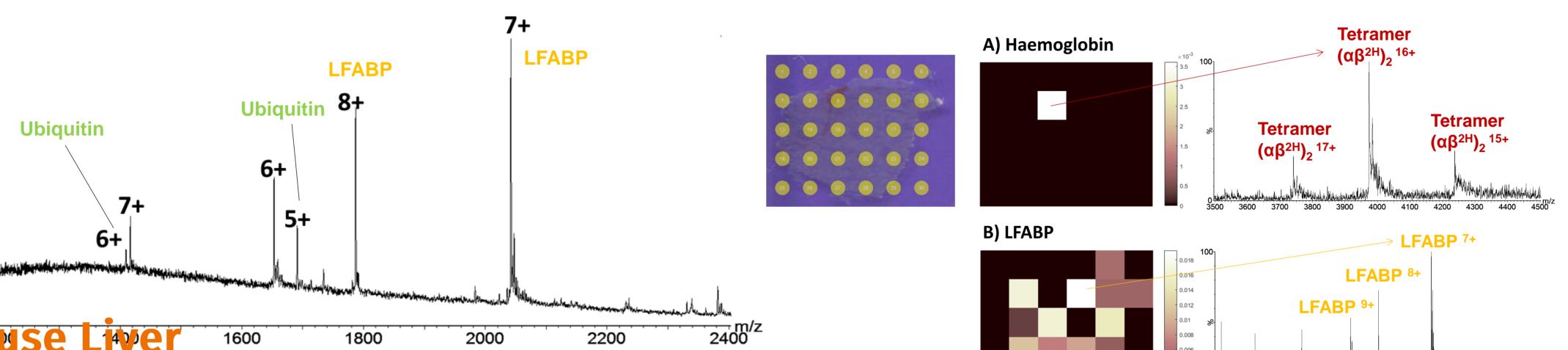
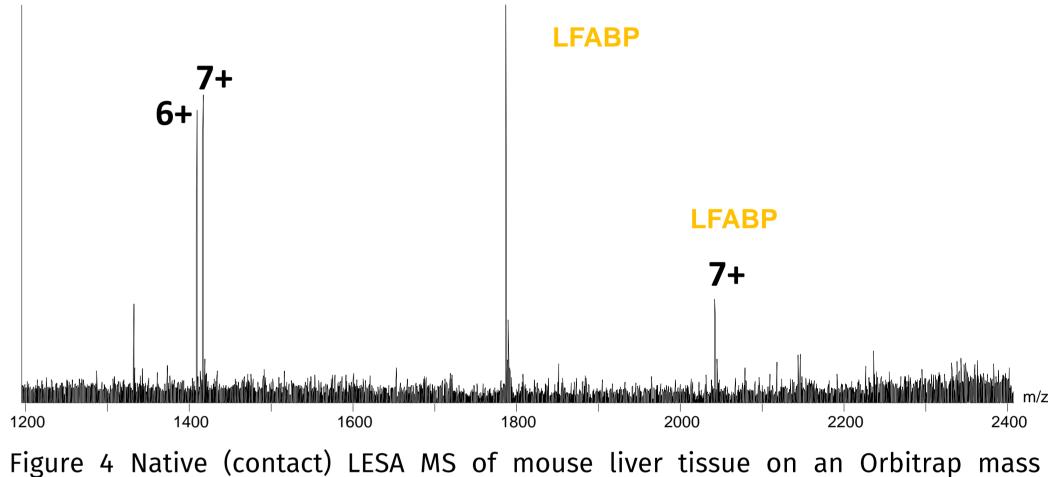


Figure 3 Native LESA MS imaging of mouse liver Sampling in locations led to the detection of predominantly LFABP in the bulk tissue and haemoglobin in a visible vasculature feature.



HRMS native LESA

spectrometer, led to the detection of abundant protein species such as liver fatty acid binding protein (LFABP) and ubiquitin in low charge states.

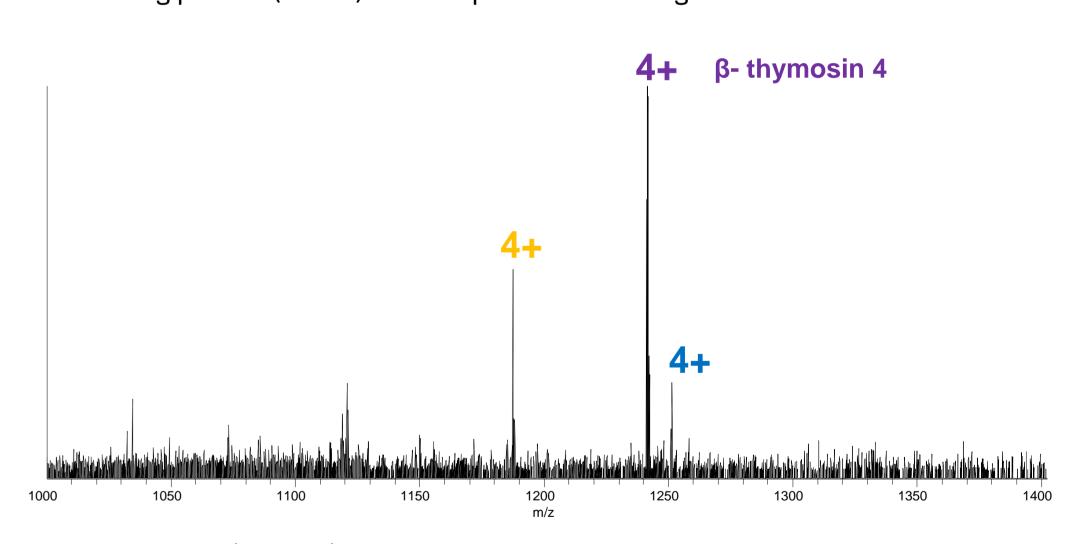


Figure 7 Native (contact) LESA MS of mouse brain tissue on an Orbitrap mass spectrometer, led to the detection of abundant protein species such as β - thymosin 4 in low charge states. Fewer protein species were detected in comparison to similar experiments on a Synapt G2S mass spectrometer as fewer gas pressures can be

Figure 2 Native (contact) LESA MS of mouse liver tissue on a Synapt G2S mass spectrometer, led to the detection of a range of protein species in the range 8-14 kDa. Species such as liver fatty acid binding protein (LFABP) and ubiquitin were detected in low charge states, indicating that the protein remains folded.

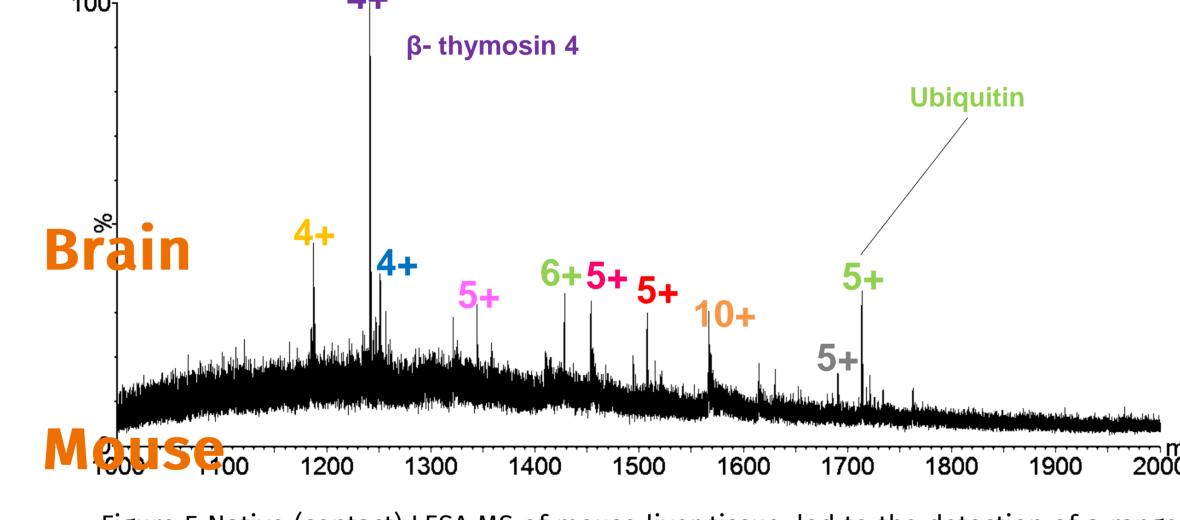
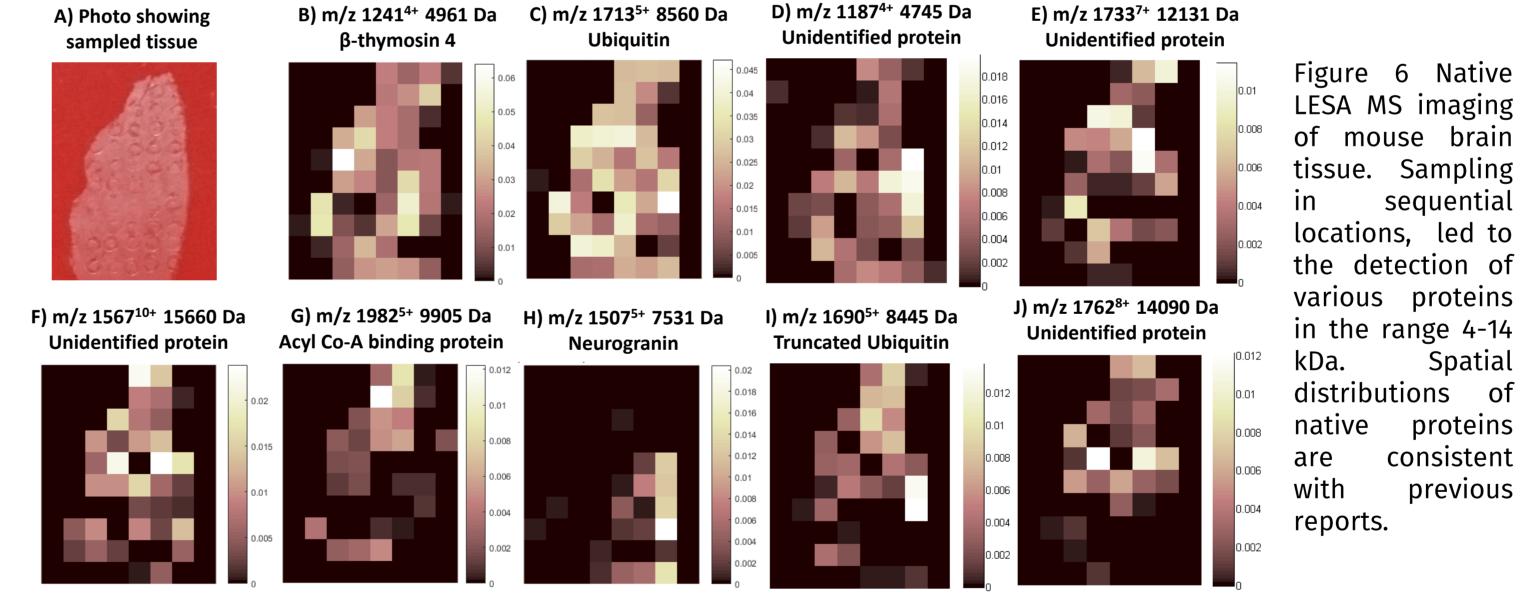
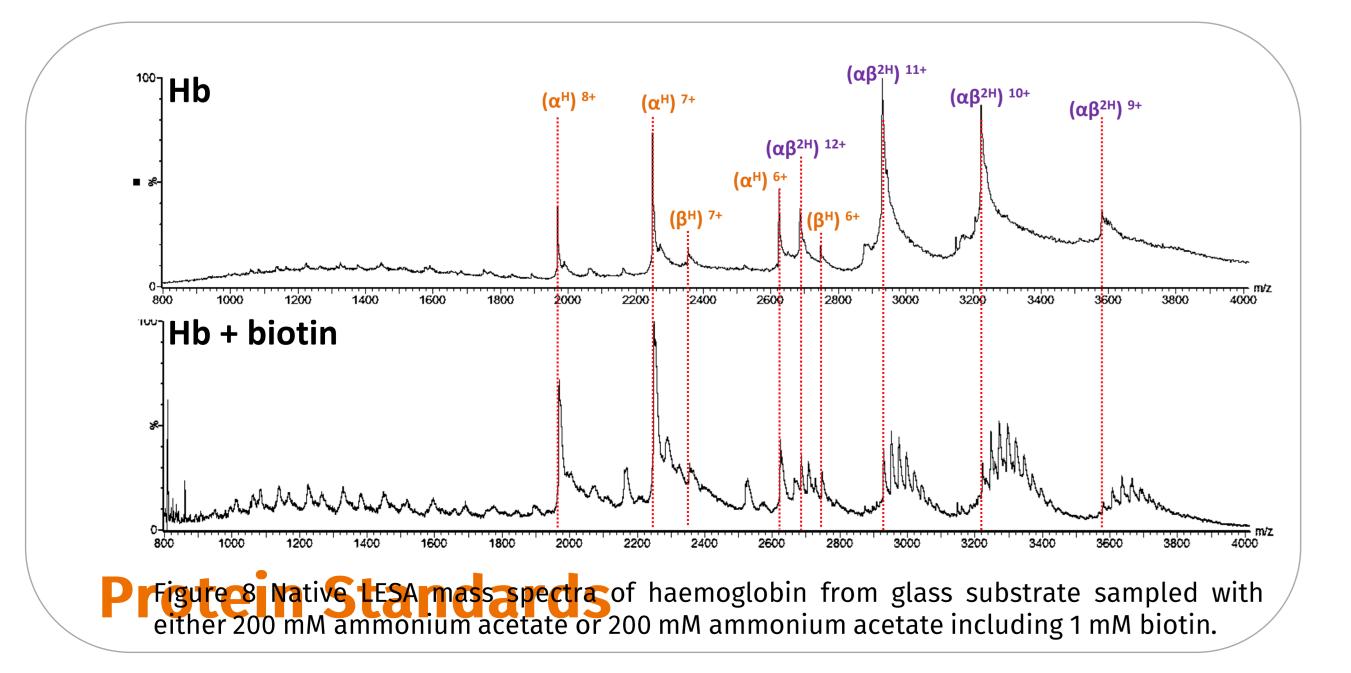


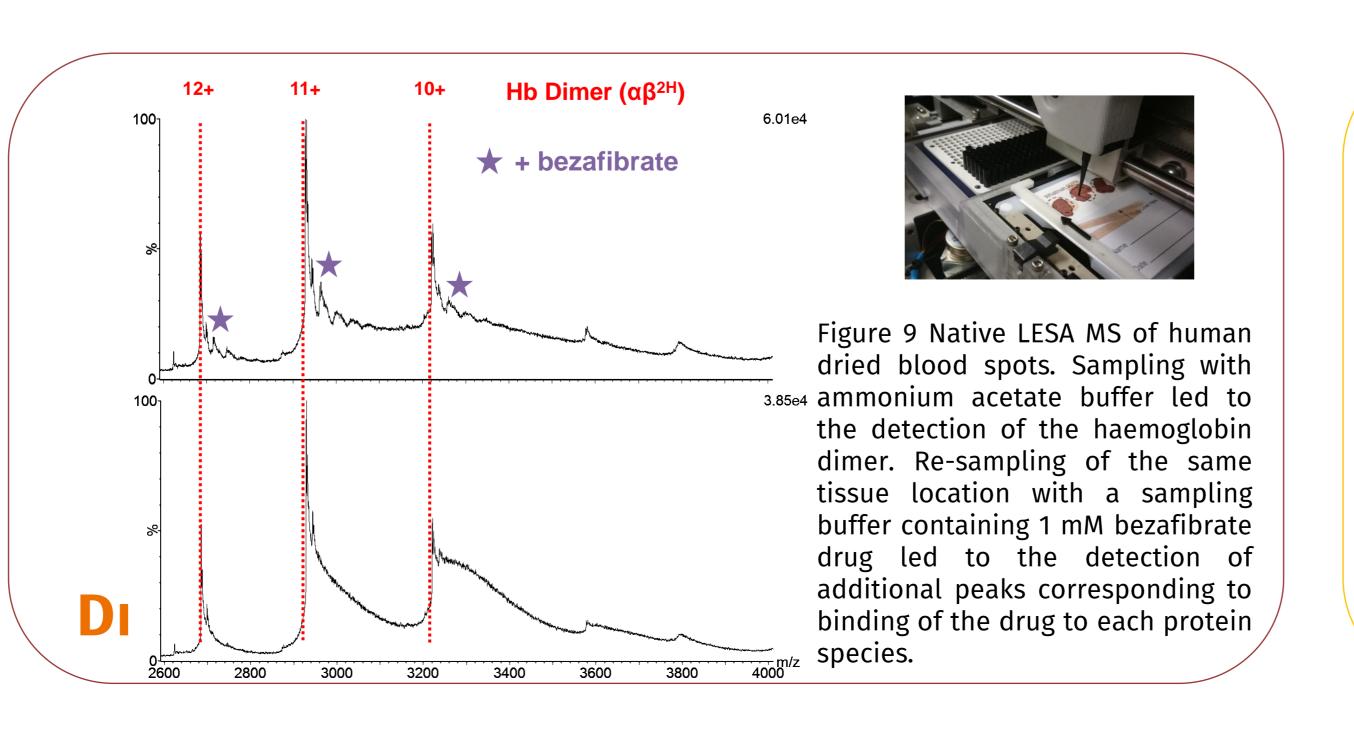
Figure 5 Native (contact) LESA MS of mouse liver tissue, led to the detection of a range of protein species in the range 4-15 kDa. Species such as β- thymosin 4 and ubiquitin were detected in low charge states, indicating that the protein remains folded.



Native LESA MS: Protein-Ligand binding

(189 kDa) and catenane hexadecamer (378 kDa). C) GroEL tetradecamer (803 kDa).





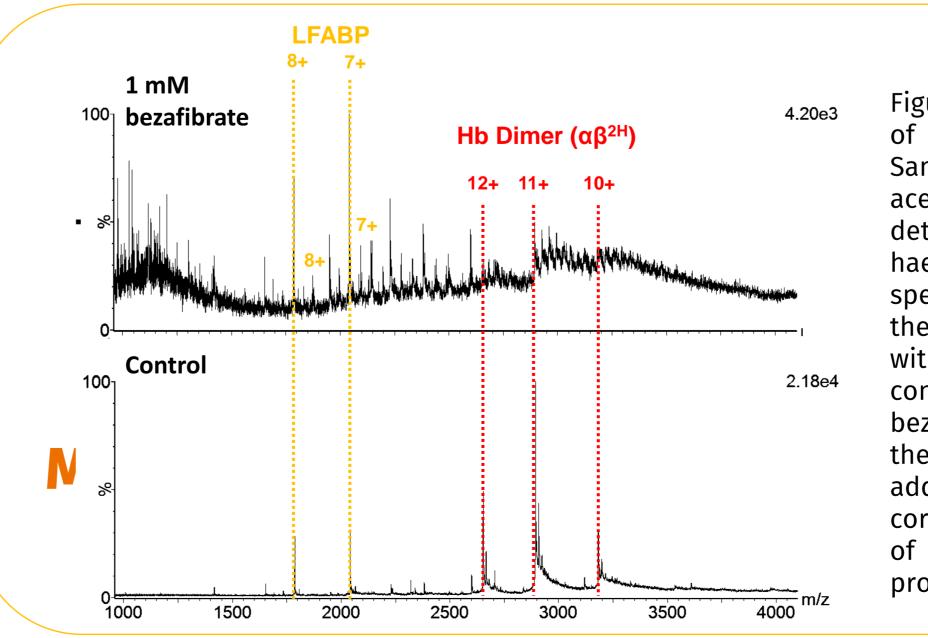


Figure 10 Native LESA MS of mouse liver tissue. Sampling with ammonium acetate buffer led to the detection of LFABP and haemoglobin dimer species. Re-sampling of the same tissue location with a sampling buffer bezafibrate drug led to detection additional corresponding to binding of the drug to each protein species.

References

Spatial

¹Edwards, R., et al., J. Am. Soc. Mass Spectrom., 2012. **23**(11): p. 1921-1930.

²Sarsby, J., et al., J. Am. Soc. Mass Spectrom., 2014. **25**(11): p. 1953-1961.

³Randall, E.C., et al., Anal. Chem., 2014. **86**(21): p. 10504-10510.

⁴Martin, N.J., et al., J. Am. Soc. Mass Spectrom., 2015. **26**(8): p. 1-8.

⁵Griffiths, R.L. and H.J. Cooper, Anal. Chem., 2016. **88**(1): p. 606-609.

⁶Mikhailov, V.M., et al., Int. J. Mass Spectrom., 2016. DOI: 10.1016/j.ijms.2016.09.01

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